

# Aflatoxin Contamination in Animal-Derived Foods and Health Risks

Rahim Aydin<sup>1,\*</sup>

<sup>1</sup>Department of Animal Nutrition and Nutritional Diseases, Balikesir University, 10100 Balikesir, Turkey

## Abstract

Aflatoxins (AFs) B<sub>1</sub>, B<sub>2</sub>, G<sub>1</sub>, and G<sub>2</sub> are important hepatotoxic mycotoxins produced by *Aspergillus flavus*, *A. parasiticus*, and *A. nomius*. They are converted into metabolites of AFM<sub>1</sub>, AFM<sub>2</sub>, B<sub>2a</sub>, and aflatoxicol by cytochrome P450-related enzymes in the liver after digestion of the feed. These metabolites accumulating in the animal-derived food products such as eggs, milk, cheese, and honey cannot be destroyed by pasteurization or heating process and may influence public health negatively. Therefore, it is very important to prevent or limit the aflatoxin contamination in the animal feeds to decrease the risk of contamination of these metabolites in animal-derived foods.

**Corresponding author:** Rahim Aydin, Department of Animal Nutrition and Nutritional Diseases, Balikesir University, 10100 Balikesir, Turkey. Email: [rahimaydin@yahoo.com](mailto:rahimaydin@yahoo.com)

**Citation:** Rahim Aydin (2020) Aflatoxin Contamination in Animal-Derived Foods and Health Risks . International Journal of Nutrition - 5(3):26-32. <https://doi.org/10.14302/issn.2379-7835.ijn-19-3100>

**Key Words:** Animal-derived foods, aflatoxins, human health

**Received:** Nov 19, 2019

**Accepted:** May 03, 2020

**Published:** May 14, 2020

**Editor:** Fei He, University of Illinois Urbana Champaign, USA.

## Introduction

Aflatoxins are toxic secondary metabolites produced by fungi of the genus *Aspergillus*, particularly *A. flavus*, *A. parasiticus*, and *A. nomius* [1]. The name "aflatoxin" was derived from the combination of "a" for the *Aspergillus* genus and "fla" for the species *flavus* and toxin meaning poison [2]. The aflatoxin problem was first recognized in 1960, when there was a severe outbreak of a disease referred to as "Turkey 'X' Disease" in the United Kingdom where more than 100,000 turkey poults and farm animals died. The cause of the disease was reported to be attributed to Brazilian peanut meal infected with *A. flavus* [3,4]. The major aflatoxins are characterized as AFB<sub>1</sub>, AFB<sub>2</sub>, AFG<sub>1</sub>, and AFG<sub>2</sub> (based on their fluorescence under UV light, blue or green) and related chromatographic mobility during thin-layer chromatography [5]. Fungal species belonging to *A. flavus* typically produce AFB<sub>1</sub> and AFB<sub>2</sub>, whereas *A. parasiticus* produces AFG<sub>1</sub> and AFG<sub>2</sub> as well as AFB<sub>1</sub> and AFB<sub>2</sub> (Figure 1).

AFB<sub>1</sub>, the most prevalent toxin in feeds, represents the greatest toxigenic and carcinogenic threat for animals and humans [7,8]. It was reported that the toxic effects of AFB<sub>1</sub> were both dose and time-dependent [9]. The total aflatoxin content can be estimated from AFB<sub>1</sub> due to a higher correlation between AFB<sub>1</sub> and total aflatoxin contents [10]. AFB<sub>1</sub> is biotransformed by cytochrome P450-associated enzymes that generate hydroxylated metabolites such as AFM<sub>1</sub> and AFB<sub>2a</sub> in the liver [11]. Aflatoxicol (AFL) can be formed by the reduction of AFB<sub>1</sub> by an NADPH-dependent cytoplasmic enzyme present in the soluble fraction of liver homogenates [12].

### *Aflatoxin Deposition and Clearance from Animal Tissues*

Feeds contaminated with AFs were shown to result in the accumulation of the metabolites in the animal tissues including liver, adipose tissues, and animal products such as milk, meat, and eggs [13,14,15]. Those metabolites may cause potential health risks in the people because they can be carried over into the animal products. After AFs were recognized in the 1960s, the Food Drug Administration (FDA) of the USA set an action level of 30 ppb of AFs in raw or finished products [16]. In 1969, the FDA revised the action level for AFs to 20 ppb for food and feed

ingredients [16]. The FDA set an action level of 0.5 ppb of AFM<sub>1</sub> in milk [16]. It was reported that only about 1-3% of the AFB<sub>1</sub> might be converted into AFM<sub>1</sub> of the milk [17].

Previously, feeding diet supplemented with AF was reported to result in the highest level of AF in the gizzard, kidney, and liver tissues [13]. Feeding a diet including 2500 ppb AFB<sub>1</sub> for 28 days was shown to cause 4.13 ppb AFB<sub>1</sub> deposition in the laying hens' liver [18]. It was shown that the levels of AFB<sub>1</sub> in the liver and kidney of chickens were significantly higher than the levels in the eggs and breast meat [19]. Residues of AFB<sub>1</sub> were detected in the eggs of hens fed supplemental 500 µg per kg feed, at levels that ranged from 0.05 to 0.16 µg/kg [20]. Laying chicken fed diets contaminated with AFB<sub>1</sub> (3300 mg/kg) for 28 days was shown to produce eggs contaminated with AFB<sub>1</sub> [13]. Also, no aflatoxin residues were recovered from whole eggs after feeding laying chickens with aflatoxin-free diet (i.e. control diet) [13]. AFM<sub>1</sub>, a metabolite of AFB<sub>1</sub>, was reported to present in the eggs of laying hens fed AFB<sub>1</sub> contaminated feed [21]. Also, it was shown that AFM<sub>1</sub> and AFM<sub>2</sub> might be recovered in the poultry litter [22]. A study was conducted in laying hens to evaluate the effect of AFB<sub>1</sub> on the egg quality in laying hens fed diet supplemented with mannan-oligosaccharides (MOS) and showed that neither AFB<sub>1</sub> nor AFM<sub>1</sub> residues were found in the eggs of groups [18]. The same study also demonstrated that hepatic levels of AFB<sub>1</sub> were significantly lower in the group fed MOS-supplemented diet compared to the group fed MOS-excluded diet [18]. It was suggested that MOS could have an ability to adsorb and degrade AFB<sub>1</sub>, reducing gastrointestinal absorption of AFB<sub>1</sub> and its levels in tissues of laying hens. In another study, synthetic zeolite was shown to have efficacy to counteract some of the toxic effects of AFs in broiler chicks [23].

Compared to the chickens, dairy cows are less sensitive to AFs due to biodegradation by rumen microorganisms [24]. In the liver, AFB<sub>1</sub> and AFB<sub>2</sub> are metabolized into AFM<sub>1</sub> and AFM<sub>2</sub>, less toxic metabolites, using cytochrome P-450 associated enzymes [15,17,25]. AFM<sub>1</sub> in the contaminated feedstuffs may be transferred into milk as AFM<sub>1</sub> in the range of 0.3-6.3% [26].

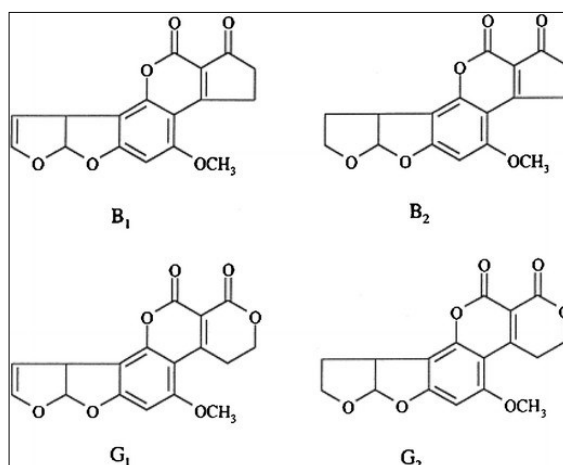


Figure 1. General structures of major aflatoxins AFB<sub>1</sub>, AFB<sub>2</sub>, AFG<sub>1</sub>, and AFG<sub>2</sub> [6].

Table 1. International legislation on AFM<sub>1</sub> in milk and dairy products for human consumption [36].

Country/region	Raw milk (µg/kg)	Dairy products (µg/kg)
Argentina	0.05	0.50 (milk products)
Austria	0.05, 0.01 (pasteurized infant milk)	0.02 (butter), 0.25 (cheese), 0.4 (powdered milk)
Brazil		0.50 (liquid milk), 5.0 powdered milk
Bulgaria	0.50	0.10 (powdered milk)
Czech Republic	0.50	
Egypt	0	0
European Union	0.05	0.05
France	0.05, 0.03 (for children <3 years)	
Honduras	0.05	0.25 (cheese)
Nigeria	1	
Rumania	0	0
Switzerland	0.05	0.025 (milk whey and products), 0.25 (cheese), 0.02 (butter)
Turkey	0.05	0.25 (cheese)
US		0.50 (liquid milk), 5.0 (powdered milk)

AFM<sub>1</sub> is very commonly detected in milk and dairy products [27,28] and concentration in the milk was shown to increase linearly depending on the level of the AFB<sub>1</sub> in the feed [29]. AFB<sub>1</sub> levels of 20% and 13.6% of the yogurt and ayran samples were found to be exceeded the maximum tolerable limit of the Turkish Food Codex [30]. Therefore, nursing animals may be affected as a result of having milk contaminated with the toxin. Those metabolites of the AFs were reported not to be destroyed during pasteurization and thermal processing [31]. A recent study showed that 36.4% of colostrum samples were found to be contaminated with an above maximum allowable level of AFB<sub>1</sub> [32]. Studies showed that milk including a significant level of AFM<sub>1</sub> may have potential risks especially for infants and children [33]. AFM<sub>1</sub> concentration in the milk was reported to decline to an undetectable level after 72 hours when the intake of AFB<sub>1</sub> is stopped [34]. Lactating cows fed a ration including 20 ppb or more AFB<sub>1</sub> was reported to produce milk that exceeds the tolerance level of the toxin in the milk.

Special attention should be paid in food for infants and young children, where more restrictive levels have been regulated. Thus, limits as low as 0.1 µg kg<sup>-1</sup> of AFB<sub>1</sub> are set for baby foods and processed cereal-based foods for infants and young children and 0.025 µg kg<sup>-1</sup> for AFM<sub>1</sub> and 0.5 µg kg<sup>-1</sup> for OTA [35]. International legislation on AFM<sub>1</sub> in milk and dairy products for human consumption is shown in Table 1.

#### *Foods Contaminated with Aflatoxins and Health Risks*

Aflatoxins are the hepatotoxic compounds causing health risks in the people consuming them more than the allowable amounts in the foods. As in the animals, these compounds or their metabolites may easily accumulate in the liver, kidney, and adipose tissues. It was reported that AFB<sub>1</sub>, the most hepatocarcinogenic compound, caused cancer mainly in the liver and other organs of animals and humans [37]. After maternal exposure of AFs during pregnancy, AFB<sub>1</sub>, AFB<sub>1</sub>-metabolites, and AFB<sub>1</sub>-albumen adducts were detected in cord blood of babies [38]. In a study conducted in Gambian children, it was reported that there was a relationship between impaired growth, particularly stunting and exposure to AFs [39,40]. The research suggested that ethnicity, dietary practice and

socio-economic status of the individuals might influence AF-exposure significantly [41].

Attempts have been made to develop methods to remove AFs from contaminated feeds or foods by physical, chemical, and biological methods [42]. It was reported that implementing advanced agricultural technologies, good agricultural, and storage practices could mitigate the mycotoxin contaminations in the products [43]. Microwave heating, treatments with ozone, or ammonia were reported to be some of the methods used for detoxification of AFs in the foods [44,45,46]. Previously, it was shown that ozone treatment could significantly reduce the level of AFs in the red pepper [47]. Recently, it was shown that AFB<sub>1</sub> could be removed by ozone treatment [48]. However, the application of ozone treatment for the degradation of AFs was reported to have limitations in food products because of the cost factor [49].

#### **Conclusions**

Chronic intake of AF-contaminated foods is a common problem especially in people of the developing countries. Contamination of crops with AFs in the field or storage may be controlled by implementing good agricultural and storage conditions. Also, identifying exposure of unacceptable AF levels in the feeds with reliable methods will decrease the exposure of AFs in the animals. Hence, minimizing exposure of domestic animals to moldy feed and taking precautions to prevent possible fungal growth in the products during the storage level will decrease AFs exposure in humans.

#### **References**

1. Moss M.O. 1998. Recent studies of mycotoxins. *Journal of Applied Microbiology*, 84:62S-76S.
2. Ellis, W.O., Smith, J.P., Simpson, B.K. 1991. Aflatoxin in food: occurrence, biosynthesis, effects on organisms, detection, and methods of control. *Critical Reviews in Food Science and Nutrition* 30: 403–439.
3. Blout, W.P. 1961. Turkey "X" disease. *Journal of the British Turkey Federation* 9, 52:55–58.
4. Asao, T., Buchi, G., Abdel-Kader, M.M., Chang, S.B., Wick, E.L., Wogan, G.N. 1963. Aflatoxins B and G. *Journal of the American Chemical Society* 85: 1706–1707.

5. Wacoo, A.P., Wendi, D., Vuzi, P.C. and Hawumba, J.F. 2014. Methods for Detection of Aflatoxins in Agricultural Food Crops. *Journal of Applied Chemistry* Article ID 706291, 15 pages.
6. Quiles, J.M., Manyes, L., Bittencourt, F., Meca, G. 2015. Effect of the oriental and yellow mustard flours as natural preservative against aflatoxins B<sub>1</sub>, B<sub>2</sub>, G<sub>1</sub> and G<sub>2</sub> production in wheat tortillas. *Journal of Food Science and Technology* 52(12): 8315–8321.
7. IARC. 1993. Some naturally occurring substances: Food items and constituents, heterocyclic aromatic amines and mycotoxins, monographs on the evaluation of carcinogenic risks to humans, vol. 56, International Agency for research on Cancer. Lyon.
8. Leeson, D.R., Diaz G.J. and Summers J.D. 1995. Poultry metabolic disorders and mycotoxins. University Books. Guelph, Canada.
9. Diaz, G.J. and Murcia, H.W. 2011. Biotransformation of aflatoxin B<sub>1</sub> and its relationship with the differential toxicological response to aflatoxin in commercial poultry species, *Aflatoxins - Biochemistry and Molecular Biology*, Dr. Ramon G. Guevara-Gonzalez (Ed.), ISBN: 978-953-307-395-8, InTech, Available from: <http://www.intechopen.com/books/aflatoxins-biochemistry-and-molecular-biology/biotransformation-of-aflatoxin-b1-and-its-relationship-with-the-differential-toxicological-response>.
10. Alptekin, Y., Duman, A. and Akkaya, M.R. 2009. Identification of fungal genus and detection of aflatoxin level in second crop corn grain. *Journal of Animal and Veterinary Advances* 8(9):1777-1779.
11. Hsieh, D. P. H. and Atkinson, D. N. 1991. Bisfuranoid mycotoxins: their genotoxicity and carcinogenicity. *Advances in Experimental Medicine and Biology*, 283: 525–532.
12. Biehl, M. L. and Buck, W. B. 1987. Chemical contaminants: their metabolism and their residues. *Journal of Food Protection*, 50: 1058–1073.
13. Wolzak A., Pearson A.M., Coleman T.H., Pestka J.J., Gray J.I. 1985. Aflatoxin deposition and clearance in the eggs of laying hens. *Food Chem. Toxicol.* 23 (12):1057-1061.
14. Pandey, I., Chauhan, S.S. 2007. Studies on production performance and toxin residues in tissues and eggs of layer chickens fed on diets with various concentrations of aflatoxin AFB<sub>1</sub>. *Br Poult Sci* 48:713–23.
15. Herzallah, S.M. 2009. Determination of aflatoxins in eggs, milk, meat and meat products using HPLC fluorescent and UV detectors. *Food Chemistry* 114: 1141–1146.
16. Price, W. D., R. A. Lovell and D. G. McChesney. 1993. Naturally occurring toxins in feedstuffs: Center for Veterinary Medicine Perspective. *Journal of Animal Science* 71(9): 2556-2562.
17. Ali N., Hashim N.H. and Yoshizawa, T. 1999. Evaluation and application of a simple and rapid method for the analysis of aflatoxins in commercial foods from Malaysia and the Philipines. *Food Additives and Contaminants* 16:273–280.
18. Zaghini, A., Martelli, G., Roncada, P., Simioli, M. and Rizzi, L. 2005. Mannanligosaccharides and aflatoxin B<sub>1</sub> in feed for laying hens: effects on egg quality, aflatoxins B<sub>1</sub> and M<sub>1</sub> residues in eggs, and aflatoxin B<sub>1</sub> levels in liver. *Poultry Sci.* 84(6): 825-832.
19. Herzallah, S.M. 2013. Aflatoxin B<sub>1</sub> residues in eggs and flesh of laying hens fed aflatoxin B<sub>1</sub> contaminated diet. *American Journal of Agricultural and Biological Sciences* 8 (2): 156-161.
20. Oliveira, C.A.F., Kobashigawa, E., Reis, T.A., Mestieri, L., Albuquerque, R. and Correa, B. 2000. Aflatoxin B<sub>1</sub> residues in eggs of laying hens fed a diet containing different levels of the mycotoxin. *Food Additives and Contaminants* 17(6): 459-462.
21. Khalil, M.M.H. Gomaa, A.M. and Sebaei, A.S. 2013. Reliable HPLC Determination of Aflatoxin M<sub>1</sub> in Eggs. *Journal of Analytical Methods in Chemistry* Volume 2013, Article ID 817091, 5 pages. <http://dx.doi.org/10.1155/2013/817091>.
22. Cortés G., Carvajal M., Méndez-Ramírez I., Avila-González E., Chilpa-Galván N., Castillo-Urueta P., Flores C.M. 2010. Identification and quantification of aflatoxins and aflatoxicol from poultry feed and their recovery in poultry litter. *Poultry Sci.* 89(5):993-1001.



23. Miazzo R., Rosa, C.A., De Queiroz, Carvalho E.C., Magnoli C., Chiacchiera S.M., Palacio, G., Saenz, M., Kikot, A., Basaldella, E., Dalcero, A. 2000. Efficacy of synthetic zeolite to reduce the toxicity of aflatoxin in broiler chicks. *Poult Sci.* 79(1):1-6.
24. Upadhaya, S.D., Park, M.A., and Ha, J.K. 2010. Mycotoxins and their biotransformation in the rumen: A review. *Asian-Aust. J. Anim. Asc.* 23(9): 1250-1260.
25. Barbieri G, Bergamini C, Ori E and Pesca P. 1994. Aflatoxin M<sub>1</sub> in parmesan cheese: HPLC determination. *Journal of Food Science* 59: 1313–1331.
26. Choudhary P. L., Sharma R. S. and Borkhateria V.N. 1998. Carry-over of aflatoxin B<sub>1</sub> from feed as aflatoxin M<sub>1</sub> in milk of Indian cows. *Milchwissenschaft* 53: 513–515.
27. Lee, J.E., Kwak, B.M., Hahn, J. and Jeon, T.H. 2009. Occurrence of aflatoxin M<sub>1</sub> in raw milk in South Korea using an immunoaffinity column and liquid chromatography. *Food Control* 20(2): 136-138.
28. Mohamadi, H. and Alizadeh, M. 2010. A Study of the Occurrence of Aflatoxin M<sub>1</sub> in dairy products marketed in Urmia, Iran. *J. Agr. Sci. Tech.* (2010) Vol. 12: 579-583.
29. Battacone G, Nudda A, Palomba M, Pascale M, Nicolussi P, Pulina G. 2005. Transfer of aflatoxin B<sub>1</sub> from feed to milk and from milk to curd and whey in dairy sheep fed artificially contaminated concentrates. *J Dairy Sci.* 88(9): 3063-3069.
30. Atasever, M.A., Atasever, M., Özturan, K. 2011. Aflatoxin M<sub>1</sub> levels in retail yoghurt and ayran in Erzurum in Turkey. *Turk J. Vet. Anim Sci.* 35(1):1-4.
31. Yitbarek, M.B., and Tamir, B. (2014). Mycotoxines and /or aflatoxins in milk and products: Review. *International Journal of Agricultural Sciences* 4(19): 294-311.
32. Radonić, J.R., KocićTanackov, S.D., Mihajlović, I.J., Grunić, Z.S., Vojinović-Miloradov, M.B., Skrinjar, M.M., Turk Sekulić, M.M. 2017. Occurrence of aflatoxin M<sub>1</sub> in human milk samples in Vojvodina, Serbia: Estimation of average daily intake by babies. *Journal of Environ Sci Health B.* 52(1):59-63.
33. Galvano, F., Galofaro, V. and Galvano, G. 1996. Occurrence and stability of aflatoxin M<sub>1</sub> in milk and milk products: A worldwide review. *J. of Food Protec.* 59: 1079-1090.
34. Van Egmond, H. 1989. Current situation on regulations of mycotoxins. Over-view of tolerances and status of standart methods of sampling and analysis. *Food Additives and Contaminants* 6: 139–188.
35. Beltran E., Ibanez M., Sancho J. V., Cortes M. A., Yusa V., Hernandez F. 2011. UHPLC-MS/MS highly sensitive determination of aflatoxins, the aflatoxin metabolite M<sub>1</sub> and ochratoxin A in baby food and milk. *Food Chem.* 126:737–744.
36. Dashti, B., S. Al-Hamli, H. Alomirah, S. Al-Zenki, A.B. Abbas, W. Sawaya. 2009. Levels of aflatoxin M<sub>1</sub> in milk, cheese consumed in Kuwait and occurrence of total aflatoxin in local and imported animal feed. *Food Control* 20: 686-690.
37. Bbosa, G.S., Kitya, D., Lubega, A., Ogwal-Okeng, J., Anokbonggo, W.W., and Kyegombe, D.B. 2013. Review of the Biological and Health Effects of Aflatoxins on Body Organs and Body Systems, Aflatoxins - Recent Advances and Future Prospects, Mehdi Razzaghi-Abyaneh, IntechOpen, DOI: 10.5772/51201. Available from: <https://www.intechopen.com/books/aflatoxins-recent-advances-and-future-prospects/review-of-the-biological-and-health-effects-of-aflatoxins-on-body-organs-and-body-systems>
38. Partanen, H.A., El-Nezami, H.S., Leppanen, J.M., Myllynen, P.K., Woodhouse, H.J., and Vahakangas, K.H. 2010. Aflatoxin B<sub>1</sub> transfer and metabolism in human placenta. *Toxicological sciences* 113(1): 216-225.
39. Turner, P.C., Moore, S.E., Hall, A.J., Prentice, A.M., Wild, C.P. 2003. Modification of immune function through exposure to dietary aflatin in Gambian children. *Environ Health Perspect* 111: 217-220.
40. Turner, P.C., Collinson, A.C., Cheung, Y.B., Gong, Y.Y., Hall, A.J., Prentice, A.M., Wild, C.P. 2007. Aflatoxin exposure in utero causes growth faltering in Gambian infants. *International Journal of Epidemiology* 36: 1119-1125.

41. Anthony, M.H., Ojochenemi, A.D., Mulunda, M., Oriyomi, S.T., Jideofor, N.F., Tunde, O., Seun, E.O., Umuhani, Y.O., Robertson, O.B., Isah, A., Halima, Y.O., Benedict, E., Umar, A, Ochai, O.D. and Aderemi, A. 2016. Aflatoxin M1 in breast milk, cow milk and milk products in minna, Nigeria and their predisposing factors. *Biochemistry and Analytical Biochemistry* 5(4): 1-6.
42. Park, D. L. 2002. Effect of processing on aflatoxin. *Advances in Experimental Medicine and Biology* 504:173–179.
43. Kamle, M., Mahato, D. K., Devi, S., Lee, K. E., Kang, S. G., and Kumar, P. 2019. Fumonisin: impact on agriculture, food, and human health and their management strategies. *Toxins* 11: 328; doi:10.3390/toxins11060328
44. Farag, R.S., Rashed, M.M., Abo-Hgger, A.A.A. 1996. Aflatoxin destruction by microwave heating. *International Journal of Food Sciences and Nutrition* 47:197–208.
45. Xu, A. 1999. Use of ozone to improve the safety of fresh fruits and vegetables. *Food Technology* 53: 58–62.
46. Prudente Jr., A.D., King, J.M. 2002. Efficacy and safety evaluation of ozonation to degrade aflatoxin in corn. *Journal of Food Science* 67: 2866–2872.
47. Inan, F., Pala, M., Doymaz, I. 2007. Use of ozone in detoxification of aflatoxin B1 in red pepper. *Journal of Stored Products Research* 43(4):425-429.
48. Agriopoulou S., Koliadima A., Karaiskakis G., Kapos J. 2016. Kinetic study of aflatoxins' degradation in the presence of ozone. *Food Control* 61:221–226.
49. Womack, E. D., Brown, A. E., and Sparks, D. L. (2014). A recent review of nonbiological remediation of aflatoxin-contaminated crops. *J. Sci. Food Agric.* 94, 1706–1714.